

Review

Therapeutic Decisions in Multiple Sclerosis Moving Beyond Efficacy

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Several innovative disease-modifying treatments (DMTs) for relapsing-remitting multiple sclerosis have been licensed recently or are in late-stage development. The molecular targets of several of these DMTs are well defined. All affect at least 1 of 4 properties, namely (1) trafficking, (2) survival, (3) function, or (4) proliferation. In contrast to β -interferons and glatiramer acetate, the first-generation DMTs, several newer therapies are imbued with safety issues, which may be attributed to their structure or metabolism. In addition to efficacy, understanding the relationship between the mechanism of action of the DMTs and their safety profile is pertinent for decision making and patient care. In this article, we focus primarily on the safety of DMTs in the context of understanding their pharmacological characteristics, including molecular targets, mechanism of action, chemical structure, and metabolism. While understanding mechanisms underlying DMT toxicities is incomplete, it is important to further develop this knowledge to minimize risk to patients and to ensure future therapies have the most advantageous benefit-risk profiles. Recognizing the individual classes of DMTs described here may be valuable when considering use of such agents sequentially or possibly in combination.

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Multiple sclerosis (MS) is a chronic central nervous system (CNS) inflammatory demyelinating disease,¹ involving both genetic and environmental factors. Its pathology is characterized by focal white and gray matter lesions with myelin, oligodendrocyte, and neuroaxonal loss²; the latter is thought to be responsible for irreversible accumulation of disability.³

There is excitement in MS therapeutics as new disease-modifying treatments (DMTs) are rapidly becoming available. However, some of this enthusiasm is tempered by risks engendered by certain newer agents. To optimally manage patients who may use these DMTs, it is important to understand and relate the DMTs' mechanisms of action (MOAs) to benefits and potential safety risks. The first DMTs, interferon beta and glatiramer acetate, reduce risk of new attacks and are generally well tolerated and safe. While activity of these agents was originally attributed to their influence on T cells, it now appears these drugs also influence innate immunity.^{4,5} Although potentially more effective and convenient, recent DMTs have been associated with risks of potentially serious adverse events (AEs), altering risk to benefit ratios. Consequently, treatment decisions have become more complex and require detailed information regarding drug properties.⁶ For many reasons, understanding risks associated with novel treatments is imperfect: (1) data collected during preclinical development are limited and extrapolation from animal to humans can be unreliable; (2) clinical studies often recruit insufficient patients to detect less common AEs, recruit highly selected patients, and may be too short to detect AEs that appear only after prolonged exposure; and (3) identifying causal re-

lationships between treatment and an AE may be difficult. Safety issues are identified after approval for around one-quarter of pharmaceutical treatments.⁷

While AEs often represent unwanted pharmacological responses related to MOA, some are idiosyncratic.⁸ Some newer therapies (eg, rituximab, alemtuzumab, BG-12, teriflunomide) also represent repurposing or modifications of previous treatments used in other diseases. Identifying and understanding AEs observed with other members of the same class, or with use of the same drug in other populations, can provide clues regarding safety and toxicities.

Herein, the safety profile of DMTs for MS is reviewed from the perspective of their molecular targets, chemical structure, MOA, and metabolism. As first-generation therapies glatiramer acetate and interferon beta present few, well-defined safety issues that have been described previously,⁹ these medications are not discussed. Instead, we focus on more recently approved therapies and those in late-stage clinical development. They are grouped into 4 categories based on their presumed target or MOA: (1) immune cell trafficking; (2) cell depletion; (3) immune cell function; and (4) cell replication (Table). Better understanding of these properties should assist physicians when choosing such therapies.

Search Strategy and Selection Criteria

References for this review were identified through searches of PubMed with the following key words: drug name (chemical and

brand name), mode of action, specific adverse effects, major metabolites, and clinical trials. The search was initially conducted August 14, 2012. Articles were also identified through searches of the authors' own files. Only articles published in English were reviewed. The final reference list was generated on the basis of originality and relevance to the broad scope of this review.

DMTs Inhibiting Immune Cell Trafficking

Acute focal CNS inflammation is triggered, particularly at early stages of disease, by influx of activated lymphocytes across the blood-brain barrier. Two types of treatment that impede lymphocyte migration have been developed and are currently licensed. These treatments prevent activated immune cells from crossing the blood-brain barrier into the CNS (natalizumab) or from exiting lymph nodes into the circulation (fingolimod). While these therapies may offer substantial efficacy, as a consequence of their MOAs they alter lymphocyte distribution, which may influence immune surveillance.

Natalizumab

Natalizumab is a humanized monoclonal antibody (mAb) (Figure 1) that has demonstrated robust reductions in clinical and radiological outcomes in relapsing-remitting MS (RRMS).^{10,11} Natalizumab is directed against the $\alpha 4$ subunit of the cell adhesion molecule very

late antigen 4 (VLA-4) expressed on the surface of lymphocytes and monocytes. Binding of VLA-4 to its receptor, vascular cell adhesion molecule 1 (VCAM-1), on vascular endothelium is required for transmigration of immune cells across the blood-brain barrier. Binding of $\alpha 4$ integrin is also required for immune cell transmigration into the gut. After successful testing in Crohn disease, natalizumab was approved for treatment of this condition.¹²

Due to blockade of leukocyte migration from blood, natalizumab treatment leads to mild leukocyte elevation¹³ and concomitant lymphocyte reduction in cerebrospinal fluid.¹⁴ On treatment discontinuation, the cerebrospinal fluid lymphocyte population reconstitutes within 6 to 12 months.¹⁴

The principal safety issue with use of natalizumab is the increased risk of progressive multifocal leukoencephalopathy (PML),¹⁵ which can be fatal or result in permanent disability. The risk of PML became evident shortly after approval of natalizumab. Two patients in the SENTINEL trial,^{15,16} which tested addition of natalizumab to weekly intramuscular interferon beta, developed PML after 28 and 37 infusions. These observations underscored the need to evaluate treatments for sufficiently long durations and for carefully designed phase 4 trials. In this regard, measuring duration of therapy may be more relevant than simply reporting "patient-years" of exposure.

The incidence of PML for patients with MS treated for at least 2 years is 5.05 in 1000 (as of February 2013).¹⁷ It may result from reactivation of JC virus within the CNS or possibly mobilization of peripheral viral reserves to the CNS.^{18,19} Three risk factors are recognized for development of PML: evidence of prior JC virus exposure, duration of natalizumab exposure, and previous use of immunosuppressants.¹⁸ A test to detect serum anti-JC virus antibodies was recently developed and serves as a useful biomarker for risk stratification in natalizumab treatment. This test should be repeated in JC virus-negative patients every 6 months owing to the annual 1% to 2% seroconversion rate.²⁰ Similarly, a high incidence of PML (1 in 500 patients) was reported with efalizumab, which was developed for treatment of psoriasis but later withdrawn. Efalizumab is an mAb directed against the adhesion molecule CD11a on T and B cells, which binds to intercellular adhesion molecule 1 (ICAM-1).²¹ Thus, this markedly elevated PML risk likely represents a class effect of these selective adhesion molecule inhibitors.

Table. Categories of Disease-Modifying Treatments for Multiple Sclerosis

Purpose	Disease-Modifying Treatment
Inhibit immune cell trafficking	Natalizumab
	Fingolimod
Promote immune cell depletion	Alemtuzumab
	Rituximab
	Ocrelizumab
Influence immune cell function	BG-12
	Laquinimod
	Daclizumab
Inhibit cell replication	Mitoxantrone
	Teriflunomide

Figure 1. Classes of Therapeutic Antibodies

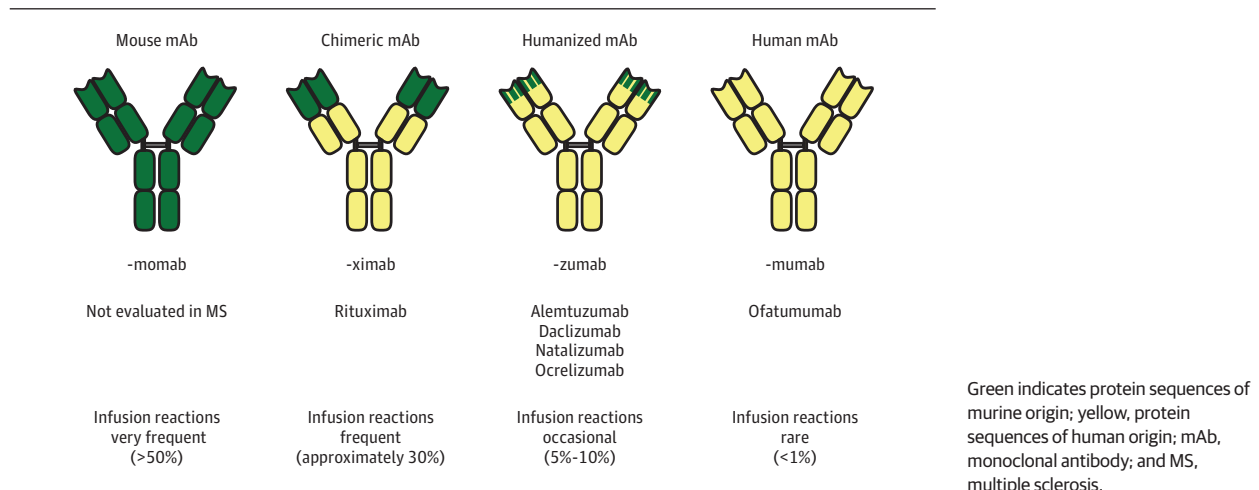
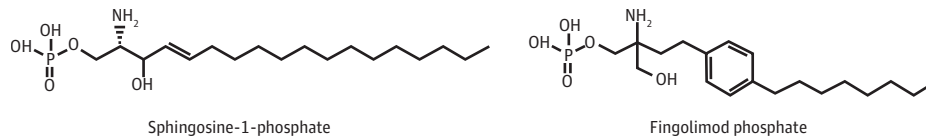


Figure 2. Chemical Structure of Sphingosine-1-Phosphate and Fingolimod



Chimeric and humanized antibodies contain murine sequences (Figure 1), which increase their immunogenicity. Use of mAbs can be associated with infusion reactions and persistent neutralizing antibodies; for natalizumab, neutralizing antibodies are associated with loss of therapeutic response and increased risk of hypersensitivity reactions.²² Recrudescence of disease activity occurs approximately 3 to 5 months after natalizumab discontinuation and corresponds to desaturation of VLA-4 binding. In some cases, natalizumab discontinuation has been associated with a rebound (overshoot) beyond baseline activity and was fatal in 1 case.²³⁻²⁵ Unfortunately, predisposing risk factors for rebound after natalizumab withdrawal have not been identified.

Fingolimod

Fingolimod (Figure 2) is an oral medication approved for treatment of RRMS. It has demonstrated superior activity to intramuscular interferon beta-1a.^{26,27} Fingolimod is a sphingosine-1-phosphate (SIP) agonist that binds to 4 of the 5 members of the SIP receptor family (SIP1, SIP2, SIP3, and SIP5). However, following binding and activation of SIP1 receptors, fingolimod acts as a functional antagonist and prevents C-C chemokine receptor type 7 (CCR7)-positive lymphocytes, including naive and central memory T cells, from exiting lymph nodes.²⁸ Consequently, lymphopenia occurs within hours of administration. Because SIP receptors are present on both neurons and glia and fingolimod penetrates the CNS,²⁹ fingolimod may exert direct CNS effects.²⁸

Few opportunistic infections have been documented in fingolimod-treated patients. Two deaths occurred from viral infections during phase 3 trials testing fingolimod, one from herpes simplex virus encephalitis and one from disseminated varicella-zoster virus, although both patients were treated with a higher dose (1.25 mg) than was approved (0.5 mg). Since approval, there has been 1 reported case of varicella-zoster virus encephalitis at 0.5 mg.³⁰ Currently, a trial is under way to determine whether 0.25 mg may be efficacious and pose less risk of viral infection.³¹ Varicella-zoster virus vaccination is recommended for patients with no history of chickenpox or prior vaccination.³² Viral infections associated with use of fingolimod are presumably linked to lymphopenia from lymphocyte sequestration. Persistent lymphopenia after drug withdrawal has been observed³³ and may also pose concern when considering initiation of another therapy soon after fingolimod discontinuation. Subtypes of SIP receptors are found in other tissues and may contribute to AEs associated with fingolimod, notably bradycardia, dyspnea, and macular edema. For example, SIP3 receptors are found in cardiac smooth muscle, vascular endothelium, and airways.³⁴ More selective SIP1 agonists are under development with the aim of eliminating certain AEs such as macular edema, a consequence from binding retinal SIP2 receptors.³⁵⁻³⁷

DMTs Producing Immune Cell Depletion

While attention has focused primarily on the role of T cells in MS pathogenesis, recent successes using B-cell-depleting agents have provided greater appreciation of the importance of this lymphocyte subset. Several mAbs originally developed for treatment of hematological malignancies, targeting B and T cells or B cells alone, are being evaluated for potential use in MS. These antibodies are IgG1 and cause cell depletion.

Alemtuzumab

Alemtuzumab, a humanized mAb (Figure 1) originally developed for treatment of B-cell chronic lymphocytic leukemia, demonstrated dramatic and sustained reductions in relapses and magnetic resonance imaging markers of disease activity in a phase 2 study³⁸ and in phase 3 studies^{39,40} vs high-dose interferon.

Alemtuzumab is directed against CD52, a surface glycoprotein present on several mature leukocyte subpopulations, including T, B, and natural killer cells.³⁸ Binding of alemtuzumab to these leukocytes leads to elimination via complement and antibody-dependent cellular cytotoxicity. However, reconstitution of leukocyte subpopulations varies⁴¹; B cells recover in approximately 6 months, whereas T cells require more than 1 year.

Treatment-induced humoral autoimmunity is a major concern associated with alemtuzumab. Graves disease, idiopathic thrombocytopenic purpura, and Goodpasture syndrome have been observed following treatment and may be life threatening without appropriate clinical management. Graves disease is the most common iatrogenic autoimmunity and occurs in up to one-quarter of alemtuzumab-treated patients,^{38,42,43} most frequently arising 12 to 18 months after starting treatment.⁴³ These humoral autoimmune disorders may relate to differences in reconstitution dynamics of B and T cells. Development of autoimmunity may also be driven by interleukin 21 (IL-21).⁴⁴ Besides autoimmunity, alemtuzumab-treated patients experienced significantly higher infection rates.

Rituximab and Ocrelizumab

Rituximab and ocrelizumab have shown robust reduction in MS disease activity in phase 2 MS trials.^{45,46} Rituximab is a chimeric mAb (Figure 1) approved for treatment of B-cell lymphoma and rheumatoid arthritis (RA).⁴⁷ Ocrelizumab is a humanized mAb. Rituximab and ocrelizumab are directed against CD20, a glycoprotein primarily found on B cells, with the exception of early progenitor (pro-B) cells and plasma cells. Binding of rituximab and ocrelizumab leads to rapid B-cell elimination that persists for 6 to 8 months without significant IgG reduction. Reduced MS activity has been attributed

to loss of B-cell-mediated cellular immunity, namely B-cell antigen presentation.^{48,49}

Severe infections have been observed in patients with lymphoma receiving rituximab. Further, development of ocrelizumab in RA and lupus was discontinued owing to occurrence of fatal opportunistic infections.⁵⁰ In addition, PML has occurred in a small number of patients with RA or lupus treated with rituximab⁵¹ and in rituximab-treated patients with lymphoma.⁵² So far, no PML cases have been associated with rituximab or ocrelizumab treatment in MS, where these agents are tested in monotherapy.

DMTs Targeting Immune Cell Function

The DMTs targeting immune cell function, or *immunomodulators*, correspond to treatments that primarily influence functional characteristics of both innate and adaptive immunity. They may affect multiple signaling pathways that alter cytokine production or effector cell functions, or both. This class includes 2 small molecules, dimethyl fumarate (DMF), BG-12, and laquinimod, and an mAb, daclizumab. A preparation of DMF, BG-12, was recently approved and laquinimod is in late-stage development. BG-12 and laquinimod may have direct central effects due to passive entry into the CNS.

BG-12

BG-12, an oral treatment, has demonstrated efficacy in 2 phase 3 RRMS trials.^{53,54} BG-12 was developed from the fumaric acid ester preparation Fumaderm, containing a mixture of DMF and monomethyl fumarate, used for psoriasis treatment in Germany. BG-12 contains only DMF and is rapidly converted to monomethyl fumarate (MMF).⁵⁵

Dimethyl fumarate and MMF activate the antioxidant transcription factor nuclear factor (erythroid-derived 2)-related factor 2 (Nrf2) pathway,^{56,57} leading to expression of detoxifying enzymes, glutathione S-transferase A2 (GSTA2), heme oxygenase 1 (HO-1), and reduced nicotinamide adenine dinucleotide phosphate (NADPH) quinone oxidoreductase 1 (NQO1).⁵⁸ Fumarates, which are electrophilic, conjugate to glutathione^{59,60} and can covalently link to essential thiol groups (nucleophiles) on macromolecules, including Keap1 (Figure 3A), the inhibitor of the Nrf2 pathway.^{57,62,63} Thus, DMF and its metabolite, MMF, activate the Nrf2 pathway by "inhibiting the inhibitor" (Figure 3B and C).

Dimethyl fumarate preserves neurons and glial cells in experimental autoimmune encephalomyelitis, while MMF protects murine neurons and human astrocytes from oxidative insult in vitro.⁵⁷ In contrast, others have reported a neuroprotective effect in vitro with DMF but not with MMF.⁶⁴ Treatment of mice with DMF induces anti-inflammatory type II dendritic cells,⁵⁶ which drive anti-inflammatory T-cell polarization.⁵⁶ Similar effects have been observed with MMF.^{56,65} Dimethyl fumarate has antiproliferative effects.⁶⁶ While potential neuroprotective effects of DMF are attributed to Nrf2 activation, whether its anti-inflammatory and immunomodulatory properties are dependent on triggering Nrf2 is unknown. In contrast, some animal studies suggest that DMF may promote renal tubular hyperplasia and oncogenic activity, also possibly related to Nrf2 activation.⁶⁷

Safety data are available from 2 BG-12 phase 3 RRMS clinical trials^{53,54} and their combined extension study.⁶⁸ Its AEs included

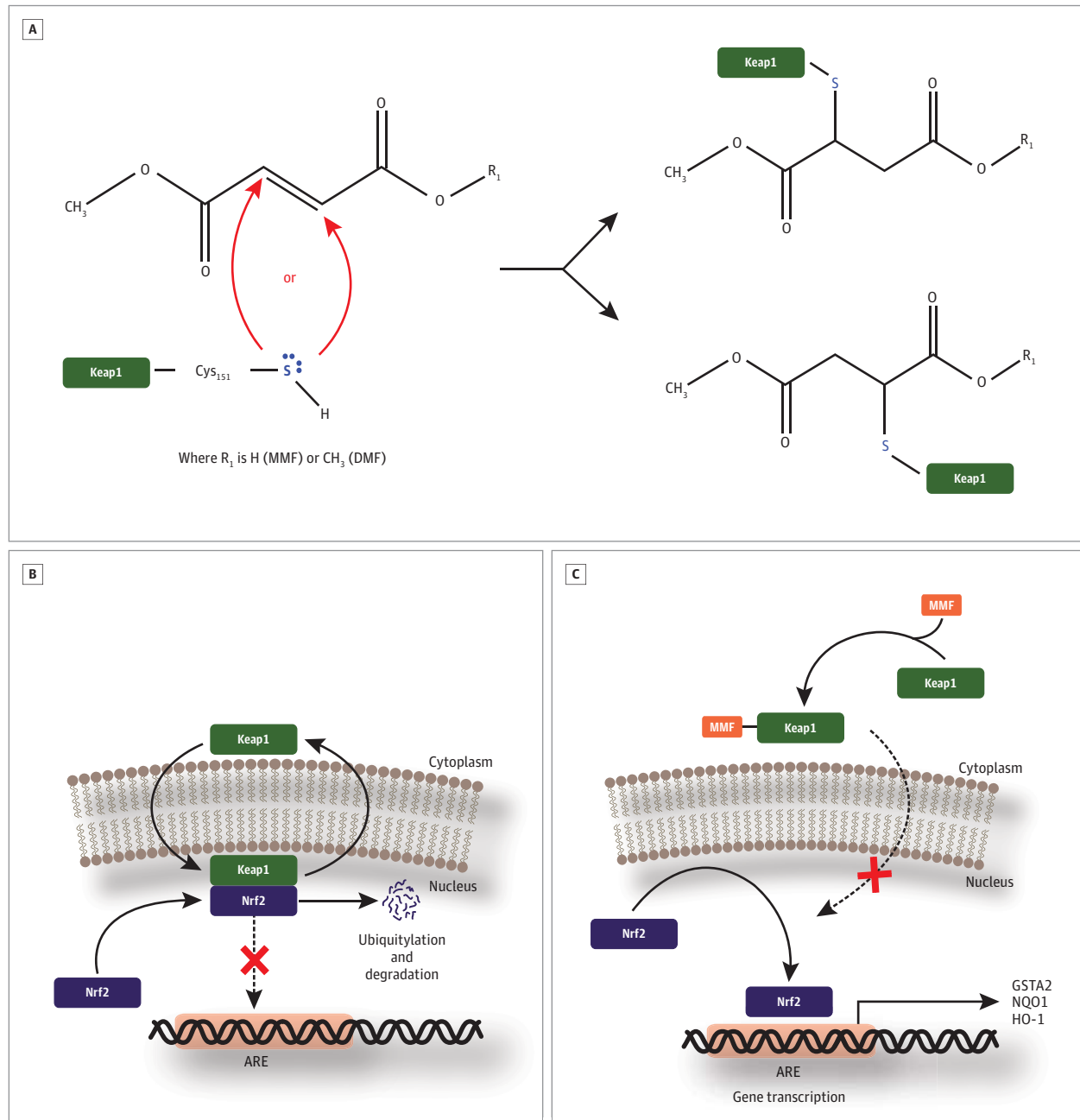
flushing, diarrhea, nausea, upper abdominal pain, decreased lymphocyte counts, and elevated liver aminotransferases.^{53,54} Renal AEs ranged from 4% to 14% and proteinuria ($\leq 5\%$) was the most common.⁶⁸ Lymphopenia was observed in 4% to 5% of BG-12-treated patients vs less than 1% in the placebo group.⁶⁸ Although no opportunistic infections were reported in the BG-12 phase 3 trials, several PML cases have been reported using fumaric acid esters in psoriasis, including 2 cases using fumaric acid ester monotherapy where PML was associated with lymphopenia that developed after initiating fumaric acid ester treatment.⁶⁹⁻⁷¹ Thus, it may be important for physicians to monitor lymphocyte counts when treating patients who have MS with BG-12.

Some of these AEs may relate to the MOA of DMF and/or its metabolites, which may be increased at higher doses.^{53,54} Following administration, DMF undergoes rapid hydrolysis to MMF and methanol.^{57,72} Interestingly, abdominal pain is a common symptom associated with methanol exposure.^{73,74} Further metabolism of MMF occurs through the tricarboxylic acid cycle, without involvement of cytochrome P450.⁶⁷ Exhalation of carbon dioxide is the primary route of elimination, accounting for approximately 60% of the DMF dose.⁶⁷ Drug-protein (eg, Keap1) adducts⁷⁵ may be responsible for liver enzyme elevations that have been reported for BG-12.⁵⁴ Flushing is thought to be attributed to release of prostaglandins causing local vasodilation.⁷⁶ Recently, bardoxolone methyl, an Nrf2 activator, was being advanced for treatment of chronic diabetic nephropathy.⁷⁷ However, its development was halted owing to deaths in the phase 3 trial testing its efficacy. Whether bardoxolone methyl toxicity is related to its activation of Nrf2, its structure, or its metabolites is not clear.

Laquinimod

Laquinimod, a quinolone-3-carboxamide, is an orally active immunomodulator that appears to have more pronounced beneficial effects on disease progression and brain atrophy than on clinical and radiological markers of inflammation in RRMS.^{78,79} Laquinimod is derived from linomide (Figure 4), whose development in MS was abandoned after occurrence of fatal serositis and myocardial infarction.⁸⁰ In evaluation of the structure-activity relationship, quinolone-3-carboxamide compounds (>60) were designed, synthesized, and evaluated in MS models.⁸¹ Individual modifications to the quinolone ring or carboxamide affected efficacy and safety, respectively. Laquinimod, containing 1 modification in the quinolone and 1 in the carboxamide, exhibited the best safety and efficacy profile⁸¹ and has since been developed for treatment of MS, Crohn disease, and lupus. Laquinimod affects the peripheral immune system and acts within the CNS. Its targets include innate immune cells, including monocytes and dendritic cells, which function as antigen-presenting cells. In experimental autoimmune encephalomyelitis, laquinimod induces anti-inflammatory antigen-presenting cells, which then downregulate proinflammatory Th1 and Th17 T cells and promote development of regulatory T cells.⁸² Glial cells, including astrocytes and microglia, are CNS targets. Laquinimod treatment reduced CNS invasion of inflammatory monocytes and prevented demyelination and subsequent axonal loss in rodents by downregulating NF- κ B signaling as well as proinflammatory cytokine and nitric oxide production in astrocytes.⁸³⁻⁸⁵ Laquinimod treatment of patients with MS was associated with elevation of brain-derived neurotrophic factor.⁸⁶

Figure 3. Methylfumarates Promote Activation of the Nrf2 Pathway via Regulation of Keap1, the Nrf2 Inhibitor



A, Methylfumarates are electrophiles that covalently bind the nucleophilic thiol group (-S-H) of Keap1 residue Cys₁₅₁.⁵⁷ Two products can be generated depending on which carbon of the π bond is conjugated. DMF indicates dimethyl fumarate; MMF, monomethyl fumarate. B, In the absence of MMF,

Keap1 binds Nrf2, promoting its ubiquitylation and consequent degradation.⁶¹ ARE indicates antioxidant response element. C, On covalent binding of MMF to Keap1, interaction between Keap1 and Nrf2 is disrupted. This stabilizes Nrf2, which permits it to bind the ARE and promote gene transcription.

Laquinimod was studied in 2 phase 3 trials using annualized relapse rate reduction as its primary end point. Because of its more pronounced beneficial effect on disability progression, a third trial is being conducted using disability as its primary end point. Safety data from the first phase 3 trials demonstrated that laquinimod was well tolerated and not associated with serious AEs; notably, serositis and myocardial infarction were not observed. Laquinimod undergoes slow hepatic metabolism, which may correlate with tran-

sient transaminase elevation seen in 5% of laquinimod-treated patients compared with 2% in placebo-treated patients.

Daclizumab

Daclizumab is a humanized nondepleting IgG1 mAb that demonstrated promising results in small pilot MS studies^{87,88} and in a phase 2 trial testing addition of daclizumab to interferon beta.⁸⁹ Two phase 2b-3 studies are under way to evaluate clinical end points.⁹⁰

Daclizumab is directed against the high-affinity α subunit (CD25) of the IL-2 receptor, which is expressed on activated T cells. Interestingly, daclizumab does not block T-cell proliferation.⁹¹ Instead, beneficial clinical and radiological measures during MS treatment were associated with expansion of regulatory CD56⁺ (bright) natural killer cells.⁹¹ No specific AEs emerged from addition of daclizumab to interferon beta,⁸⁹ although liver enzyme elevations and cutaneous reactions were observed.

DMTs Targeting Immune Cell Replication

The recognized role of lymphocytes in MS pathogenesis has provided the foundation for advancing drugs that inhibit their expansion. In this class, mitoxantrone hydrochloride and teriflunomide are agents approved for MS treatment that target DNA.

Mitoxantrone

Mitoxantrone is an anthracenedione approved for treatment of rapidly evolving relapsing or secondary progressive MS.⁹² It is an anti-neoplastic agent used for treatment of metastatic breast cancer, acute myeloid leukemia and non-Hodgkin lymphoma.

Mitoxantrone is an inhibitor of topoisomerase II⁹³ and can intercalate into double-stranded DNA. Mitoxantrone affects all proliferating cells and is therefore nonselective, although it appears to inhibit B cells more than T cells. Like the related anthracycline chemotherapeutics, mitoxantrone is associated with dose-dependent cardiotoxic effects.⁹⁴ Initially, the recognized risk of therapy-related acute leukemia in MS treatment was 0.25%, but 10 years after mitoxantrone approval, this risk approached 1.0%.⁹⁴ This increased risk of therapy-related acute leukemia provides another

example underscoring the importance of vigilant postapproval safety monitoring. Because of concerns for cardiotoxic effects and therapy-related acute leukemia, use of mitoxantrone for MS is generally confined to second- or third-line treatment.

Teriflunomide

Teriflunomide is an oral agent that demonstrated efficacy in phase 3 clinical trials for treatment of RRMS⁹⁵ and was recently approved in the United States. Teriflunomide is the active metabolite of leflunomide (Figure 5), a DMT licensed for treatment of RA.⁹⁶ Teriflunomide inhibits mitochondrial dihydroorotate dehydrogenase, an enzyme used for de novo synthesis of pyrimidine nucleotides in proliferating cells. However, teriflunomide does not inhibit the salvage pathway used by resting cells.⁹⁷

The AEs associated with teriflunomide include lymphopenia, alopecia, elevated liver enzymes, elevated blood pressure, and nausea. Leflunomide and teriflunomide are considered to be teratogenic in humans and are therefore contraindicated in pregnancy.⁹⁸ Teriflunomide can also penetrate into breast milk.⁹⁹ As leflunomide treatment of RA is associated with elevated risk of tuberculosis, purified protein derivative testing is recommended before commencing teriflunomide treatment in patients with MS.⁹⁹

Teriflunomide undergoes extensive enterohepatic recirculation, leading to long-term exposure of the liver to high concentrations that may result in hepatotoxic effects,¹⁰⁰ an important safety issue with leflunomide in RA^{101,102} and teriflunomide in MS.⁹⁹ As a consequence of its enterohepatic recycling, substantial time is required to achieve steady-state plasma concentrations of teriflunomide. The extended 10-day half-life¹⁰⁰ is of potential clinical relevance in case of serious AE or pregnancy, when rapid drug elimination is necessary. In this context, wash-out procedures have

Figure 4. Chemical Structure of Linomide and Laquinimod

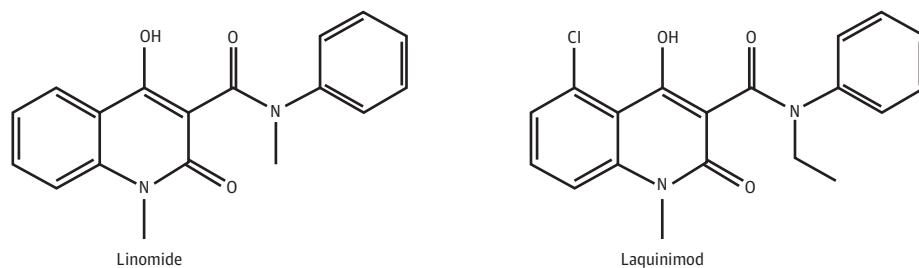
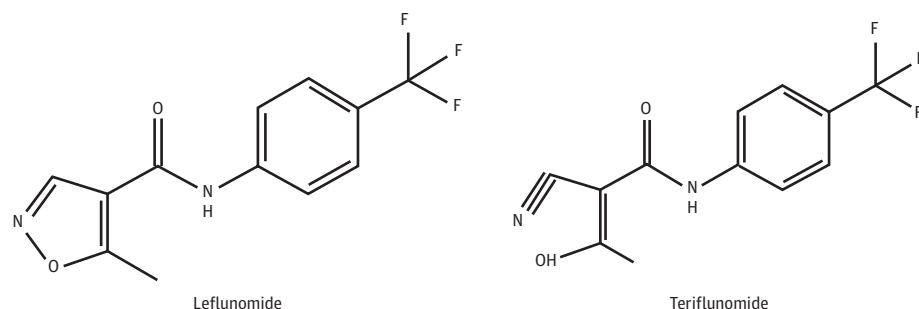


Figure 5. Chemical Structure of Leflunomide and Teriflunomide



been developed involving administration of cholestyramine or activated charcoal to prevent enterohepatic recirculation. Although genetic polymorphisms of cytochrome P450 isoforms have been associated with AEs from leflunomide,¹⁰³ cytochrome P450 may have a limited role in teriflunomide metabolism.

Discussion

With introduction of several new MS medications, treatment decisions are becoming more complex. Whereas efficacy remains paramount, choosing new agents necessitates careful consideration of other characteristics, including MOA, duration of effect (ie, pharmacodynamics), and potential risks. In this article, we have classified DMTs into 4 categories based on their ability to (1) inhibit cell trafficking, (2) promote immune cell depletion, (3) influence immune function, or (4) inhibit cell replication. While we have provided a framework, it is important to recognize that each category is not mutually exclusive. Agents that reduce lymphocyte proliferation may induce immune modulation and vice versa.^{66,104} Nevertheless, categorization of agents with similarities can help us anticipate specific AEs of newer agents. In this regard, it is important to recognize that natalizumab and efalizumab, which are selective adhesion molecule inhibitors and therefore block lymphocyte trafficking, are both associated with PML. While newer S1P agonists (eg, BAF312 and ONO-4641) selectively activate S1P1 receptors on lymphocytes and reduce trafficking, these agents also bind the S1P1 receptors expressed by cells directing atrioventricular conduction and therefore, like fingolimod, can be associated with some level of bradycardia. A potential hazard of DMTs like mitoxantrone and teriflunomide, which interact with DNA or inhibit DNA metabolism, respectively, is that such agents may affect DNA within stem cells and/or germ cells.^{94,99}

Agents specific for one molecular target or immune pathway may have pleiotropic effects. While the intended mechanism of a given DMT may shift immune balance favorably for one disease, it may have paradoxical activity in others. Tumor necrosis factor receptor antagonists are widely used for RA and were considered for MS therapy until their use was associated with increased risk of CNS demyelination. Although T- and B-cell depletion by alemtuzumab is associated with potent therapeutic effects in MS, its use promotes humoral autoimmunity targeting the thyroid and, more rarely, platelets, kidney, or lung. Whether this iatrogenic autoimmunity relates to distinct kinetics of T- and B-cell reconstitution or abnormal T-cell cytokine secretion is not clear. Prolonged lymphopenia after alemtuzumab treatment may be an important consideration when using other agents sequentially. Specifically, should one wait until there is full reconstitution of both B cells and T cells prior to treatment with another agent? Similarly, if a patient does not respond to fingolimod, one may consider delaying sequential treatment until the fingolimod-associated lymphopenia resolves. Interestingly, prolonged lymphopenia and associated immunosuppression, rather than concern related to its clinical benefit in MS, probably halted development and use of cladribine. When treating MS with newer agents, we may need to think beyond our next therapy.

Physicians who treat MS will need to pay particular attention to metabolic properties when prescribing certain newer agents. In contrast to interferons (natural endogenous proteins) and glatiramer acetate (a polypeptide-based agent), newer oral therapies are synthetic organic molecules and may be metabolized and excreted differently. Teriflunomide undergoes prolonged hepatobiliary circulation; in certain situations (eg, pregnancy or AE) it may be necessary to accelerate teriflunomide elimination. Metabolites may be active therapeutically and also responsible for adverse effects. Dimethyl fumarate is rapidly metabolized to MMF, considered the predominant bioactive form responsible for Nrf2 activation. As methanol is produced in metabolism of DMF to MMF, methanol or other DMF metabolites could possibly contribute to its adverse effects.

With introduction of new agents that use different MOAs, one can envisage combining MS medications that may act in an additive or synergistic manner.¹⁰⁵ Although this is a worthy goal, there are practical concerns. First, to establish that 2 effective drugs are more efficacious together than either one alone may require enrolling large numbers of patients. Second, as the price of many MS agents increases, it may be unreasonable to consider the added cost in combination. In general, one should be cautious when combining pharmacological agents as their metabolism may interfere with one another and further paradoxical effects can occur. In this regard, clinical trials have suggested that widely used cholesterol-lowering statins may interfere with the efficacy of interferon beta,^{106,107} and it is postulated that this potential antagonistic effect relates to their opposing activity on the proinflammatory signaling molecule STAT1.¹⁰⁸

Surrogate markers that associate risk of adverse effects, or response, to DMTs are particularly helpful in clinical practice. As JC virus antibody-positive patients have increased risk of PML during natalizumab treatment, anti-JC virus seropositivity has become an important biomarker for stratification of this risk. Serum IL-21 levels could be considered to estimate risk of thyroid autoimmunity in alemtuzumab-treated patients. Stratification may include gene polymorphisms. For example, ABC-transporter gene polymorphisms have been associated with response to mitoxantrone.¹⁰⁹

In stark contrast to the excitement surrounding our increasing repertoire of treatments for RRMS, the paucity of useful agents for progressive MS is sobering. Thus far, our successes primarily target the peripheral inflammation characterizing RRMS, but not the CNS-resident inflammatory and neurodegenerative processes of progressive MS. Hopefully, this therapeutic gap will be breached through better understanding of MS progression, refining our clinical and imaging metrics of MS progression, and testing established and novel agents with potential antioxidative and neuroprotective MOAs.

While no drug to date cures MS, it is clear that major advances have been made in therapeutics of RRMS. However, several current drugs have serious, sometimes life-threatening toxic effects. Although the understanding of mechanisms underlying DMT toxicities is incomplete, it is important to develop this knowledge to minimize risk to patients and to ensure that future therapies have the most advantageous benefit to risk profiles. Recognizing the individual classifications of DMTs described here may be beneficial when considering use of such agents sequentially or eventually in combination.

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REFERENCES

- Koch-Henriksen N, Sørensen PS. The changing demographic pattern of multiple sclerosis epidemiology. *Lancet Neurol*. 2010;9(5):520-532.
- Compston A, Coles A. Multiple sclerosis. *Lancet*. 2008;372(9648):1502-1517.
- Brück W, Stadelmann C. Inflammation and degeneration in multiple sclerosis. *Neurol Sci*. 2003;24(suppl 5):S265-S267.
- Prod'homme T, Zamvil SS. Bench to bedside: tempering antigen-presenting cells in multiple sclerosis. *Nat Med*. 2008;14(6):614-615.
- Weber MS, Prod'homme T, Youssef S, et al. Type II monocytes modulate T cell-mediated central nervous system autoimmune disease. *Nat Med*. 2007;13(8):935-943.
- Kieseier BC, Stüve O. A critical appraisal of treatment decisions in multiple sclerosis: old vs new. *Nat Rev Neurol*. 2011;7(5):255-262.
- Lexchin J. New drugs and safety: what happened to new active substances approved in Canada between 1995 and 2010? *Arch Intern Med*. 2012;172(21):1680-1681.
- Rawlins MD, Thomson JW. Mechanisms of adverse drug reactions. In: Davies DM, ed. *Textbook of Adverse Drug Reactions*. 4th ed. New York, NY: Oxford University Press; 1991.
- Wingerchuk DM. Multiple sclerosis disease-modifying therapies: adverse effect surveillance and management. *Expert Rev Neurother*. 2006;6(3):333-346.
- Polman CH, O'Connor PW, Havrdova E, et al; AFFIRM Investigators. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med*. 2006;354(9):899-910.
- Rudick RA, Stuart WH, Calabresi PA, et al; SENTINEL Investigators. Natalizumab plus interferon beta-1a for relapsing multiple sclerosis. *N Engl J Med*. 2006;354(9):911-923.
- Sandborn WJ, Yednock TA. Novel approaches to treating inflammatory bowel disease: targeting alpha-4 integrin. *Am J Gastroenterol*. 2003;98(11):2372-2382.
- Krumbholz M, Meinl I, Kumpfel T, Hohlfeld R, Meinl E. Natalizumab disproportionately increases circulating pre-B and B cells in multiple sclerosis. *Neurology*. 2008;71(17):1350-1354.
- Stüve O, Marra CM, Jerome KR, et al. Immune surveillance in multiple sclerosis patients treated with natalizumab. *Ann Neurol*. 2006;59(5):743-747.
- Langer-Gould A, Atlas SW, Green AJ, Bollen AW, Pelletier D. Progressive multifocal leukoencephalopathy in a patient treated with natalizumab. *N Engl J Med*. 2005;353(4):375-381.
- Kleinschmidt-DeMasters BK, Tyler KL. Progressive multifocal leukoencephalopathy complicating treatment with natalizumab and

- interferon beta-1a for multiple sclerosis. *N Engl J Med*. 2005;353(4):369-374.
17. Global natalizumab (TYSABRI) safety update. Weston, MA: Biogen Idec; February 5, 2013.
18. Bloomgren G, Richman S, Hotermans C, et al. Risk of natalizumab-associated progressive multifocal leukoencephalopathy. *N Engl J Med*. 2012;366(20):1870-1880.
19. Monaco MC, Major EO. The link between VLA-4 and JC virus reactivation. *Expert Rev Clin Immunol*. 2012;8(1):63-72.
20. Gorelik L, Lerner M, Bixler S, et al. Anti-JC virus antibodies: implications for PML risk stratification. *Ann Neurol*. 2010;68(3):295-303.
21. Major EO. Progressive multifocal leukoencephalopathy in patients on immunomodulatory therapies. *Annu Rev Med*. 2010;61:35-47.
22. Tysabri [package insert]. Cambridge, MA: Biogen Idec; 2012.
23. Lenhard T, Biller A, Mueller W, Metz I, Schönberger J, Wildemann B. Immune reconstitution inflammatory syndrome after withdrawal of natalizumab? *Neurology*. 2010;75(9):831-833.
24. Ramos-Cejudo J, Oreja-Guevara C, Stark Aroeira L, Rodriguez de Antonio L, Chamorro B, Diez-Tejedor E. Treatment with natalizumab in relapsing-remitting multiple sclerosis patients induces changes in inflammatory mechanism. *J Clin Immunol*. 2011;31(4):623-631.
25. Rigau V, Mania A, Béfort P, et al. Lethal multiple sclerosis relapse after natalizumab withdrawal. *Neurology*. 2012;79(22):2214-2216.
26. Cohen JA, Barkhof F, Comi G, et al; TRANSFORMS Study Group. Oral fingolimod or intramuscular interferon for relapsing multiple sclerosis. *N Engl J Med*. 2010;362(5):402-415.
27. Kappos L, Radue EW, O'Connor P, et al; FREEDOMS Study Group. A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. *N Engl J Med*. 2010;362(5):387-401.
28. Cohen JA, Chun J. Mechanisms of fingolimod's efficacy and adverse effects in multiple sclerosis. *Ann Neurol*. 2011;69(5):759-777.
29. Foster CA, Howard LM, Schweitzer A, et al. Brain penetration of the oral immunomodulatory drug FTY720 and its phosphorylation in the central nervous system during experimental autoimmune encephalomyelitis: consequences for mode of action in multiple sclerosis. *J Pharmacol Exp Ther*. 2007;323(2):469-475.
30. Ratchford JN, Costello K, Reich DS, Calabresi PA. Varicella-zoster virus encephalitis and vasculopathy in a patient treated with fingolimod. *Neurology*. 2012;79(19):2002-2004.
31. MS study evaluating safety and efficacy of two doses of fingolimod versus copaxone [NCT01633112]. <http://clinicaltrials.gov/ct2/show/NCT01633112>. Accessed March 11, 2013.
32. Gilenya [package insert]. East Hanover, NJ: Novartis Pharmaceuticals; 2012.
33. Johnson TA, Shames I, Keezer M, et al. Reconstitution of circulating lymphocyte counts in FTY720-treated MS patients. *Clin Immunol*. 2010;137(1):15-20.
34. Brinkmann V, Baumruker T. Pulmonary and vascular pharmacology of sphingosine 1-phosphate. *Curr Opin Pharmacol*. 2006;6(3):244-250.
35. Olsson T, Boster A, Fernandez O, et al. Efficacy and safety of ponesimod, an oral, selective sphingosine 1-phosphate receptor-1 modulator, in patients with relapsing-remitting multiple sclerosis: results from a phase IIb, randomised, double-blind, placebo-controlled trial. *Mult Scler*. 2012;18(4)(suppl):49-50.
36. Stuve O, Selmaj K, Li D, et al. BAF312, a selective sphingosine-1-phosphate receptor modulator improves MRI and clinical outcomes in relapsing-remitting multiple sclerosis (RRMS) (S30.001). *Neurology*. 2012;78(meeting abstracts 1):S30.001. doi:10.1212/WNL.78.1_MeetingAbstracts.S30.001.
37. Vollmer T, Selmaj K, Bar-Or A, Zipp FA. Double-blind, placebo-controlled, phase 2, 26-week DreaMS trial of a selective S1P receptor agonist ONO-4641 in patients with relapsing-remitting multiple sclerosis. *Neurology*. 2012;79(11):e87-e91.
38. Coles AJ, Compston DA, Selmaj KW, et al; CAMMS223 Trial Investigators. Alemtuzumab vs interferon beta-1a in early multiple sclerosis. *N Engl J Med*. 2008;359(17):1786-1801.
39. Cohen JA, Coles AJ, Arnold DL, et al; CARE-MS I Investigators. Alemtuzumab vs interferon beta 1a as first-line treatment for patients with relapsing-remitting multiple sclerosis: a randomised controlled phase 3 trial. *Lancet*. 2012;380(9856):1819-1828.
40. Coles AJ, Twyman CL, Arnold DL, et al; CARE-MS II Investigators. Alemtuzumab for patients with relapsing multiple sclerosis after disease-modifying therapy: a randomised controlled phase 3 trial. *Lancet*. 2012;380(9856):1829-1839.
41. Coles AJ, Wing M, Smith S, et al. Pulsed monoclonal antibody treatment and autoimmune thyroid disease in multiple sclerosis. *Lancet*. 1999;354(9191):1691-1695.
42. Coles A, Brinar V, Arnold D, et al. Efficacy and Safety Results from Comparison of Alemtuzumab and Rebif Efficacy in Multiple Sclerosis I (CARE-MS I): a phase 3 study in relapsing-remitting treatment-naïve patients (S01.006). *Neurology*. 2012;78(meeting abstracts 1):S01.006. doi:10.1212/WNL.78.1_MeetingAbstracts.S01.006.
43. Cossburn M, Pace AA, Jones J, et al. Autoimmune disease after alemtuzumab treatment for multiple sclerosis in a multicenter cohort. *Neurology*. 2011;77(6):573-579.
44. Jones JL, Phuah CL, Cox AL, et al. IL-21 drives secondary autoimmunity in patients with multiple sclerosis, following therapeutic lymphocyte depletion with alemtuzumab (Campath-1H). *J Clin Invest*. 2009;119(7):2052-2061.
45. Hauser SL, Waubant E, Arnold DL, et al; HERMES Trial Group. B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. *N Engl J Med*. 2008;358(7):676-688.
46. Kappos L, Li D, Calabresi PA, et al. Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial. *Lancet*. 2011;378(9805):1779-1787.
47. Edwards JC, Szczepanski L, Szechinski J, et al. Efficacy of B-cell-targeted therapy with rituximab in patients with rheumatoid arthritis. *N Engl J Med*. 2004;350(25):2572-2581.
48. Bar-Or A, Fawaz L, Fan B, et al. Abnormal B-cell cytokine responses a trigger of T-cell-mediated disease in MS? *Ann Neurol*. 2010;67(4):452-461.
49. Weber MS, Prod'homme T, Patarroyo JC, et al. B-cell activation influences T-cell polarization and outcome of anti-CD20 B-cell depletion in central nervous system autoimmunity. *Ann Neurol*. 2010;68(3):369-383.
50. F. Hoffmann-La Roche Ltd. Roche and Biogen Idec decide to suspend Ocrelizumab treatment: rheumatoid arthritis development programme on hold. http://www.roche.com/media/media_releases/med-cor-2010-03-08.htm. Accessed March 11, 2013.
51. Palazzo E, Yahia SA. Progressive multifocal leukoencephalopathy in autoimmune diseases. *Joint Bone Spine*. 2012;79(4):351-355.
52. Rituxan [package insert]. South San Francisco, CA: Genentech; 2012.
53. Fox RJ, Miller DH, Phillips JT, et al; CONFIRM Study Investigators. Placebo-controlled phase 3 study of oral BG-12 or glatiramer in multiple sclerosis. *N Engl J Med*. 2012;367(12):1087-1097.
54. Gold R, Kappos L, Arnold DL, et al; DEFINE Study Investigators. Placebo-controlled phase 3 study of oral BG-12 for relapsing multiple sclerosis. *N Engl J Med*. 2012;367(12):1098-1107.
55. Litjens NH, Burggraaf J, van Strijen E, et al. Pharmacokinetics of oral fumarates in healthy subjects. *Br J Clin Pharmacol*. 2004;58(4):429-432.
56. Ghoreschi K, Brück J, Kellerer C, et al. Fumarates improve psoriasis and multiple sclerosis by inducing type II dendritic cells. *J Exp Med*. 2011;208(11):2291-2303.
57. Linker RA, Lee DH, Ryan S, et al. Fumaric acid esters exert neuroprotective effects in neuroinflammation via activation of the Nrf2 antioxidant pathway. *Brain*. 2011;134(pt 3):678-692.
58. Nguyen T, Nioi P, Pickett CB. The Nrf2-antioxidant response element signaling pathway and its activation by oxidative stress. *J Biol Chem*. 2009;284(20):13291-13295.
59. Rostami Yazdi M, Mrowietz U. Fumaric acid esters. *Clin Dermatol*. 2008;26(5):522-526.
60. Rostami-Yazdi M, Clement B, Schmidt TJ, Schinor D, Mrowietz U. Detection of metabolites of fumaric acid esters in human urine: implications for their mode of action. *J Invest Dermatol*. 2009;129(1):231-234.
61. Baird L, Dinkova-Kostova AT. The cytoprotective role of the Keap1-Nrf2 pathway. *Arch Toxicol*. 2011;85(4):241-272.
62. Frycák P, Zdráhal Z, Ulrichová J, Wiegrebe W, Lemr K. Evidence of covalent interaction of fumaric acid esters with sulfhydryl groups in peptides. *J Mass Spectrom*. 2005;40(10):1309-1318.
63. Schmidt TJ, Ak M, Mrowietz U. Reactivity of dimethyl fumarate and methylhydrogen fumarate towards glutathione and N-acetyl-L-cysteine: preparation of S-substituted thiosuccinic acid esters. *Bioorg Med Chem*. 2007;15(1):333-342.
64. Albrecht P, Bouchachia I, Goebels N, et al. Effects of dimethyl fumarate on neuroprotection and immunomodulation. *J Neuroinflammation*. 2012;9:163.
65. Litjens NH, Rademaker M, Ravensbergen B, Thio HB, van Dissel JT, Nibbering PH. Effects of monomethylfumarate on dendritic cell differentiation. *Br J Dermatol*. 2006;154(2):211-217.

66. Lehmann JC, Listopad JJ, Rentzsch CU, et al. Dimethylfumarate induces immunosuppression via glutathione depletion and subsequent induction of heme oxygenase 1. *J Invest Dermatol*. 2007;127(4):835-845.
67. Tecfidera [package insert]. Cambridge, MA: Biogen Idec; 2013.
68. Phillips JT, Fox RJ, Selmaj K, et al. Long-term safety and tolerability of oral BG-12 (dimethyl fumarate) in relapsing-remitting multiple sclerosis: interim results from ENDORSE. *Mult Scler*. 2012;18(4)(suppl):517-518.
69. Sweetser MT, Dawson KT, Bozic C. Manufacturer's response to case reports of PML. *N Engl J Med*. 2013;368(17):1659-1661.
70. Ermis U, Weis J, Schulz JB. PML in a patient treated with fumaric acid. *N Engl J Med*. 2013;368(17):1657-1658.
71. van Oosten BW, Killestein J, Barkhof F, Polman CH, Wattjes MP. PML in a patient treated with dimethyl fumarate from a compounding pharmacy. *N Engl J Med*. 2013;368(17):1658-1659.
72. Litjens NH, van Strijen E, van Gulpen C, et al. In vitro pharmacokinetics of anti-psoriatic fumaric acid esters. *BMC Pharmacol*. 2004;4:22.
73. Barceloux DG, Bond GR, Krenzelok EP, Cooper H, Vale JA; American Academy of Clinical Toxicology Ad Hoc Committee on the Treatment Guidelines for Methanol Poisoning. American Academy of Clinical Toxicology practice guidelines on the treatment of methanol poisoning. *J Toxicol Clin Toxicol*. 2002;40(4):415-446.
74. Kraut JA, Kurtz I. Toxic alcohol ingestions: clinical features, diagnosis, and management. *Clin J Am Soc Nephrol*. 2008;3(1):208-225.
75. Evans DC, Watt AP, Nicoll-Griffith DA, Baillie TA. Drug-protein adducts: an industry perspective on minimizing the potential for drug bioactivation in drug discovery and development. *Chem Res Toxicol*. 2004;17(1):3-16.
76. Hanson J, Gille A, Offermanns S. Role of HCA (GPR109A) in nicotinic acid and fumaric acid ester-induced effects on the skin. *Pharmacol Ther*. 2012;136(1):1-7.
77. Reisman SA, Chertow GM, Hebbar S, Vaziri ND, Ward KW, Meyer CJ. Bardoxolone methyl decreases megalin and activates nrf2 in the kidney. *J Am Soc Nephrol*. 2012;23(10):1663-1673.
78. Comi G, Jeffery D, Kappos L, et al; ALLEGRO Study Group. Placebo-controlled trial of oral laquinimod for multiple sclerosis. *N Engl J Med*. 2012;366(11):1000-1009.
79. Vollmer TL, Soelberg Sorensen P, Arnold DL; BRAVO Study Group. A placebo-controlled and active comparator phase III trial (BRAVO) for relapsing-remitting multiple sclerosis. *Mult Scler*. 2011;17(10)(suppl):S507-S508.
80. Noseworthy JH, Wolinsky JS, Lublin FD, et al; North American Linomide Investigators. Linomide in relapsing and secondary progressive MS, part I: trial design and clinical results. *Neurology*. 2000;54(9):1726-1733.
81. Jönsson S, Andersson G, Fex T, et al. Synthesis and biological evaluation of new 1,2-dihydro-4-hydroxy-2-oxo-3-quinolinecarboxamides for treatment of autoimmune disorders: structure-activity relationship. *J Med Chem*. 2004;47(8):2075-2088.
82. Schulze-Topphoff U, Shetty A, Varrin-Doyer M, et al. Laquinimod, a quinoline-3-carboxamide, induces type II myeloid cells that modulate central nervous system autoimmunity. *PLoS One*. 2012;7(3):e33797.
83. Mishra MK, Wang J, Silva C, Mack M, Yong VW. Kinetics of proinflammatory monocytes in a model of multiple sclerosis and its perturbation by laquinimod. *Am J Pathol*. 2012;181(2):642-651.
84. Brück W, Pförtner R, Pham T, et al. Reduced astrocytic NF- κ B activation by laquinimod protects from cuprizone-induced demyelination. *Acta Neuropathol*. 2012;124(3):411-424.
85. Ruffini F, Rossi S, Bergamaschi A, et al. Laquinimod prevents inflammation-induced synaptic alterations occurring in experimental autoimmune encephalomyelitis [published online December 11, 2012]. *Mult Scler*. doi:10.1177/1352458512469698.
86. Thöne J, Ellrichmann G, Seubert S, et al. Modulation of autoimmune demyelination by laquinimod via induction of brain-derived neurotrophic factor. *Am J Pathol*. 2012;180(1):267-274.
87. Rose JW, Burns JB, Bjorklund J, Klein J, Watt HE, Carlson NG. Daclizumab phase II trial in relapsing and remitting multiple sclerosis: MRI and clinical results. *Neurology*. 2007;69(8):785-789.
88. Bielekova B, Richert N, Howard T, et al. Humanized anti-CD25 (daclizumab) inhibits disease activity in multiple sclerosis patients failing to respond to interferon beta. *Proc Natl Acad Sci U S A*. 2004;101(23):8705-8708.
89. Wynn D, Kaufman M, Montalban X, et al; CHOICE Investigators. Daclizumab in active relapsing multiple sclerosis (CHOICE study): a phase 2, randomised, double-blind, placebo-controlled, add-on trial with interferon beta. *Lancet Neurol*. 2010;9(4):381-390.
90. Gold R, Giovannoni G, Selmaj K, et al; SELECT Study Investigators. Daclizumab high-yield process in relapsing-remitting multiple sclerosis (SELECT): a randomised, double-blind, placebo-controlled trial. *Lancet*. 2013;381(9884):2167-2175.
91. Bielekova B, Catalfamo M, Reichert-Scriver S, et al. Regulatory CD56(bright) natural killer cells mediate immunomodulatory effects of IL-2R α -targeted therapy (daclizumab) in multiple sclerosis. *Proc Natl Acad Sci U S A*. 2006;103(15):5941-5946.
92. Hartung HP, Gonsette R, König N, et al; Mitoxantrone in Multiple Sclerosis Study Group (MIMS). Mitoxantrone in progressive multiple sclerosis: a placebo-controlled, double-blind, randomised, multicentre trial. *Lancet*. 2002;360(9350):2018-2025.
93. Malonne H, Atassi G. DNA topoisomerase targeting drugs: mechanisms of action and perspectives. *Anticancer Drugs*. 1997;8(9):811-822.
94. Marriott JJ, Miyasaki JM, Gronseth G, O'Connor PW; Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology. Evidence report: the efficacy and safety of mitoxantrone (Novantrone) in the treatment of multiple sclerosis: report of the Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology. *Neurology*. 2010;74(18):1463-1470.
95. O'Connor P, Wolinsky JS, Confavreux C, et al; TEMSO Trial Group. Randomized trial of oral teriflunomide for relapsing multiple sclerosis. *N Engl J Med*. 2011;365(14):1293-1303.
96. Fox RI, Herrmann ML, Frangou CG, et al. Mechanism of action for leflunomide in rheumatoid arthritis. *Clin Immunol*. 1999;93(3):198-208.
97. Fairbanks LD, Bofill M, Ruckemann K, Simmonds HA. Importance of ribonucleotide availability to proliferating T-lymphocytes from healthy humans: disproportionate expansion of pyrimidine pools and contrasting effects of de novo synthesis inhibitors. *J Biol Chem*. 1995;270(50):29682-29689.
98. Fukushima R, Kanamori S, Hirashiba M, et al. Inhibiting the teratogenicity of the immunosuppressant leflunomide in mice by supplementation of exogenous uridine. *Toxicol Sci*. 2009;108(2):419-426.
99. Aubagio [package insert]. Cambridge, MA: Genzyme; 2012.
100. Rozman B. Clinical pharmacokinetics of leflunomide. *Clin Pharmacokinet*. 2002;41(6):421-430.
101. Alcorn N, Saunders S, Madhok R. Benefit-risk assessment of leflunomide: an appraisal of leflunomide in rheumatoid arthritis 10 years after licensing. *Drug Saf*. 2009;32(12):1123-1134.
102. US Food and Drug Administration. FDA Drug Safety Communication: new boxed warning for severe liver injury with arthritis drug Arava (leflunomide). <http://www.fda.gov/Drugs/DrugSafety/PostmarketDrugSafetyInformationforPatientsandProviders/ucm218679.htm>. Accessed March 11, 2013.
103. Bohanec Grabar P, Rozman B, Tomsic M, Suput D, Logar D, Dolzan V. Genetic polymorphism of CYP1A2 and the toxicity of leflunomide treatment in rheumatoid arthritis patients. *Eur J Clin Pharmacol*. 2008;64(9):871-876.
104. Claussen MC, Korn T. Immune mechanisms of new therapeutic strategies in MS: teriflunomide. *Clin Immunol*. 2012;142(1):49-56.
105. Stüve O, Youssef S, Weber MS, et al. Immunomodulatory synergy by combination of atorvastatin and glatiramer acetate in treatment of CNS autoimmunity. *J Clin Invest*. 2006;116(4):1037-1044.
106. Birnbaum G, Cree B, Altafullah I, Zinser M, Reder AT. Combining beta interferon and atorvastatin may increase disease activity in multiple sclerosis. *Neurology*. 2008;71(18):1390-1395.
107. Sorensen PS, Lycke J, Erälänpä J, et al; SIMCOMBIN Study Investigators. Simvastatin as add-on therapy to interferon β -1a for relapsing-remitting multiple sclerosis (SIMCOMBIN study): a placebo-controlled randomised phase 4 trial. *Lancet Neurol*. 2011;10(8):691-701.
108. Zamvil SS, Steinman L. Combining statins with interferon β in multiple sclerosis: think twice, it might not be all right. *Lancet Neurol*. 2011;10(8):672-673.
109. Cotte S, von Ahnen N, Kruse N, et al. ABC-transporter gene-polymorphisms are potential pharmacogenetic markers for mitoxantrone response in multiple sclerosis. *Brain*. 2009;132(pt 9):2517-2530.